

Characterization of *Arachis hypogaea* L. oil obtained from different extraction techniques and *in vitro* antioxidant potential of supercritical fluid extraction extract

Abstract

Aim: The present investigation was aimed to characterize the fixed oil of *Arachis hypogaea* L. using five different extraction methods: Supercritical fluid extraction (SFE), ultrasound assistance extraction, soxhlet extraction, solvent extraction, and three phase partitioning method. **Materials and Methods:** The SFE conditions (temperature, pressure, and volume of CO₂) were optimized prior for better yield. The extracted oils were analyzed and compared for their physiochemical parameters, high-performance thin layer chromatography (HPTLC), gas chromatography-mass spectrometry (GC-MS), and Fourier transform infrared spectrometry (FT-IR) fingerprinting. Anti-oxidant activity was also determined using DPPH and superoxide scavenging method. **Results:** The main fatty acids were oleic, linoleic, palmitic, and stearic acids as obtained by GC-MS. HPTLC analysis revealed the presence of similar major components in chromatograms. Similarly, the pattern of peaks as obtained in FT-IR and GC-MS spectra of same oils by different extraction methods was superimposable. **Conclusion:** Analysis reported that the fixed oil of *A. hypogaea* L. is a good source of unsaturated fatty acid, mainly n-6 and n-9 fatty acid with a significant antioxidant activity of oil obtained from SFE extraction method.

Key words:

Fourier transforms infrared spectroscopy, gas chromatography-mass spectrometry, supercritical fluid extraction

Introduction

Peanut (*Arachis hypogaea* L.) is an important oilseed in the world and an important food source of lipids and proteins. Peanut is composed of about 50% lipid and 29% of protein and also contain vitamins and minerals.^[1] Peanut oil is one of the major oils in the human diet and rich in unsaturated fatty acids (80%),^[2] to which it has attributed its effectiveness in reducing total cholesterol. Because of its beta-sitosterol, it may inhibit cancer growth.^[3]

Conventional industrial processing of peanut oil involves pressing and solvent (SOL) extraction. Pressing is less efficient and gives lower yield (40–60%). SOL extraction, although the recovery of oil is in the 90–98% range, has

disadvantages including high investment and energy requirements. The hexane used as the most common solvent for extraction of oil is listed among hazardous air pollutants

Rishika Chauhan, Iftexhar Ahmad, Yasmeen Khan,
Ennus Tajuddin Tamboli, Sayeed Ahmad

Department of Pharmacognosy and Phytochemistry, Faculty of
Pharmacy, Jamia Hamdard, New Delhi, India

Address for correspondence:

Dr. Sayeed Ahmad,
Department of Pharmacognosy and Phytochemistry, Faculty of Pharmacy,
Jamia Hamdard, New Delhi - 110 062, India.
E-mail: sahmaj_h@jamiyahamdard.ac.in

This is an open access article distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 3.0 License, which allows others to remix, tweak, and build upon the work non-commercially, as long as the author is credited and the new creations are licensed under the identical terms.

For reprints contact: reprints@medknow.com

How to cite this article: Chauhan R, Ahmad I, Khan Y, Tamboli ET, Ahmad S. Characterization of *Arachis hypogaea* L. oil obtained from different extraction techniques and *in vitro* antioxidant potential of supercritical fluid extraction extract. Drug Dev Ther 2016;7:87-91.

Access this article online	
Website: www.ddtjournal.org	Quick Response Code 
DOI: 10.4103/2394-6555.191150	

associated with neurological and respiratory disorders on prolonged exposure (the International Standard Organization permits only 50 ppm residual hexane in oilseed meal).^[4] Hence, there is a need of another technique that is safe, efficient, and less time-consuming. Supercritical fluid extraction (SFE) has emerged as an attractive technique for the extraction of fixed oils. Supercritical CO₂ is a greener alternative to the hexane in lipid extraction because it is less toxic and eliminates solvent waste. In addition, energy costs associated with reaching the supercritical state for CO₂ have been shown to be less than energy costs associated with conventional solvent distillation.

In this study, we have characterized fixed oil of peanuts extracted by SFE and other conventional methods. Along with it, we also studied the antioxidant potential of extracted fixed oil of peanuts.

Materials and Methods

Plant materials

Peanut was collected from a local market and the same was identified and authenticated by a pharmacognosist and voucher specimen was deposited in the Department of Pharmacognosy and Phytochemistry, Jamia Hamdard, New Delhi (Specimen No. BNPL/JH/RCMPH: 05/2012). The peanuts were cleaned, dried, grinded, and passed through a sieve (40 mesh).

Extraction of fixed oil of peanut

Soxhlet (SOXH) and ultrasound (US)-assisted extraction was done using hexane (ratio 1:3) as a solvent, for 4 h at boiling point in SOXH extraction whereas 1 h at 54 ± 2°C with occasional stirring in US.^[5] Fixed oil was obtained by three-phase partitioning (TPP) method as per the reported method.^[6] SFE extraction was performed at a pressure of 400 bar and a temperature of 50°C for 75 min at CO₂ flow rate of 20 g/min,^[7] which was optimized before extraction.

Physiochemical parameters

Physiochemical parameters (acid value [AV], peroxide value [PV], and iodine value [IV]), ester value, saponification value, and unsaponification value were analyzed according to Indian Pharmacopoeia 2007^[8] and matched with reported values.

High-performance thin layer chromatography profiling

All the five samples obtained through five different extraction procedures (50 times diluted) were applied in duplicate (5.0 µL each) on precoated silica gel 60 F₂₅₄ plates (E. Merck, 0.20 mm thickness) using Linomat V. The chromatograms were scanned at 300 nm after development. The plate was also scanned at 370 nm after spraying with ethanolic sulfuric acid, 10% v/v and drying at 100°C for 5 min in oven.

Gas chromatography-mass spectrometry analysis of fatty acids

The fixed oils extracted using different extraction methods were analyzed for their fatty acid composition by preparing fatty acid methyl ester.^[9]

Agilent 7890A series, gas chromatography-mass spectrometry system was used for analysis attached with CTC-PAL, HP-5 ms automatic sampler, and mass detector. The split less mode at 250°C inlet temperature, 0.1 ml injection volume with helium gas at 1 ml/min, and 70–202°C of oven temperature was used for analysis.

Fourier transform infrared spectrometry analysis

For the analysis, a Fourier transform-infrared spectrometry spectrophotometer (IRAffinity, Shimadzu) equipped with a deuterated triglycine sulfate detector with a resolution of 4 cm⁻¹ was used. The data interval provided by the instrument for a resolution of 4 cm⁻¹ is 1.93 cm⁻¹. All spectra were recorded from 4000 to 400 cm⁻¹. A thin film of the fixed oil was created between two polished KBr disks.^[10]

Assessment of *in vitro* antioxidant activity

Free radical scavenging activity was determined by using DPPH assay previously reported by Hasan *et al.*^[11] and Liu *et al.*^[12] The method described by Liu *et al.*^[13] was used for the determination of anion scavenging activity.

Results and Discussion

Evaluation of extraction methods

Table 1 represents the yields of extraction for peanut oil. The result revealed that SFE technique achieved the best result (41.80%) as the extraction was carried out at 400 bars. It was previously reported that oil yield increases with an increase in the pressure that lead to an increase in CO₂ density, resulting in a higher solubility and hence higher extraction yield.

Physiochemical parameter

The physiochemical parameters of the extracted oil of peanut were compared [Table 2]. The extracted oil using SFE technique showed lowest AV (0.52 ± 0.7 mg KOH/g) indicating less free fatty acid. The PV of SFE oil was lowest indicating less rancidity. The IV was highest in TPP showing more unsaturation in extracted oil. The saponification value was lower in SOXH followed by SFE which indicates that the weight of fatty acid was larger in triglycerides.

High-performance thin layer chromatography profiling

The best separation of constituents of oil samples was found in the solvent system hexane: Diethyl ether: Acetic acid (7:4:1, v/v/v). The comparative high-performance thin layer chromatography fingerprinting of oils extracted was found matching with the presence of different common

compounds at matching R_f [Figure 1]. R_f 0.63 and R_f 0.57 were the major components of all five peanut oil extracted through different methods at 300 nm and 370 nm, respectively. The triglycerides, major components of the fixed oil having high affinity to the solvent system while complex lipids such as phospholipids having low affinity remained at the origin of chromatogram.

Table 1: Percentage yield of extracted oil of peanut extracted by five extraction methods

Extraction procedure	Percentage yield of extracted oil (% w/w)
Solvent	33.68
Soxhlet	29.88
Ultrasound-assisted	29.00
Three-phase partitioning	40.67
Supercritical fluid extraction	41.80

Gas chromatography-mass spectrometry analysis

The fatty acid compositions of the extracted oil of peanut are shown in Table 3. Oleic acid, linoleic acid, palmitic acid, and stearic acids were found to be the major fatty acids in the five extracted samples. The oil consists of two classes of polyunsaturated fatty acid, n-9 (oleic acid) and n-6 (linoleic acid) at R_t 23.00 min and 22.00 min, respectively [Figure 2]. The oils by SOL extraction showing the highest percent of area are oleic acid and linoleic acid.

Fourier transform infrared spectrometry fingerprinting of extracted fixed oil of peanut

The intense peaks in the spectra were two intensive bands at 2926 cm^{-1} (aliphatic CH_2 asymmetric) and 2856 cm^{-1} (symmetric stretching vibration); at 1741 cm^{-1} is assigned to the C=O stretching vibration of the ester carbonyl functional group of the triglycerides; at 1455 cm^{-1} is observed a band

Table 2: Physiochemical parameters of the extracted fixed oil of peanut

Physiochemical parameter	Extraction method				
	SOL*	SOXH*	US*	TPP*	SFE*
Acid value (mg KOH/g oil)	0.61 ± 0.8	0.59 ± 0.6	0.60 ± 0.5	0.63 ± 0.5	0.52 ± 0.2
Ester value (mg KOH/g Oil)	302.49 ± 0.7	293.62 ± 0.9	297.06 ± 0.6	301.43 ± 1.2	295.42 ± 0.4
Iodine value	48.03 ± 0.6	46.93 ± 1.0	47.29 ± 0.8	49.16 ± 0.4	47.85 ± 0.6
Peroxide value (mEq of active oxygen/kg oil)	3.38 ± 1.5	4.73 ± 0.3	4.19 ± 1.3	2.58 ± 0.7	2.49 ± 0.8
Saponification value (mg KOH/g oil)	303.10 ± 0.6	294.21 ± 0.9	297.66 ± 1.0	302.06 ± 1.4	295.98 ± 1.0
Unsaponification value (g/kg)	2.16 ± 1.1	2.36 ± 0.8	2.59 ± 0.6	1.63 ± 0.4	2.29 ± 0.7

*SFE – supercritical fluid extraction; TPP – Three-phase partitioning; SOXH – Soxhlet; US – Ultrasound; SOL – Solvent

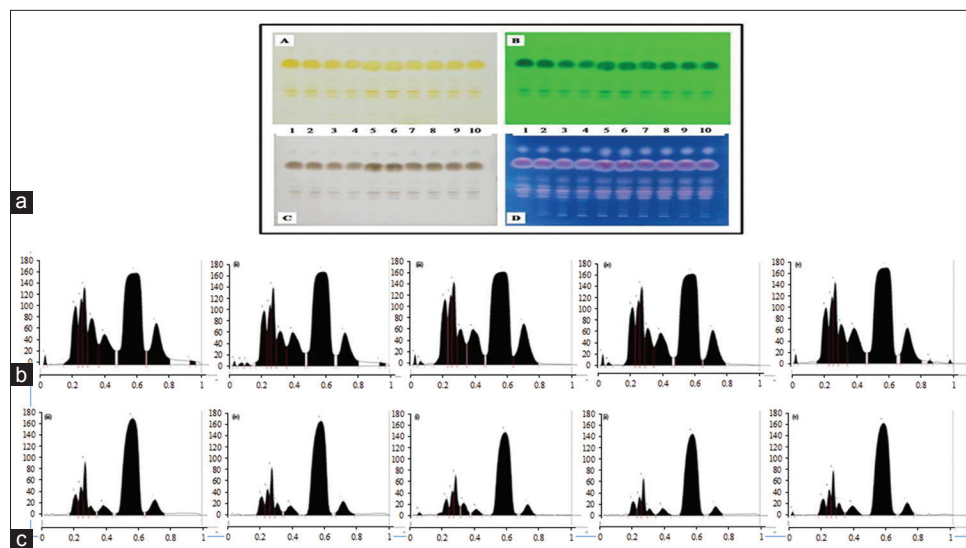


Figure 1: (a) High-performance thin layer chromatography fingerprint of peanut oil; derivatization: Iodine vapor A: Day light, B: 254 nm; 10% ethanolic sulfuric acid C: Day light, D: 366 nm; supercritical fluid extraction: Track 1–2, three-phase partitioning: 3–4, soxhlet: 5–6, ultrasound: 7–8, solvent: 9–10; (b) high-performance thin layer chromatography chromatograms (iodine derivatized) at 300 nm; (c) High-performance thin layer chromatography chromatograms (10% ethanolic sulfuric acid derivatized) at 370 nm; (i) supercritical fluid extraction, (ii) three-phase partitioning, (iii) soxhlet, (iv) ultrasound-assisted extraction, (v) solvent extraction

which is assigned to C = H scissors deformation vibration; the bands at 1165 cm⁻¹ and 1236 cm⁻¹ are assigned to the

vibration of the C-O ester groups and CH₂ group; the band near 1371 cm⁻¹ is assigned to the bending vibration of CH₂ groups; at 719 cm⁻¹ (overlapping of CH₂ rocking), 3007 cm⁻¹ (C-H stretching vibration of the cis-double) [Figure 3]. There were some weak peaks at 590 cm⁻¹ (-CH₂ rocking vibration) and 3474 cm⁻¹ (overtone of the glyceride ester carbonyl).

Table 3: Fixed oil content of fixed oil peanut

Fatty acid	Percentage of area				
	SOL*	SOXH*	US*	TPP*	SFE*
Palmitic acid	12.94	11.90	12.62	12.82	12.65
Steric acid	4.53	4.26	4.37	4.83	4.30
Linoleic acid	32.48	29.6	31.46	30.2	32.21
Oleic acid	43.18	40.11	42.79	40.98	40.73

*SFE – Supercritical fluid extraction; TPP – Three phase partitioning; SOXH – Soxhlet; US – Ultrasound; SOL – Solvent

In vitro antioxidant activity of supercritical fluid extraction-extracted fixed oil

The peanut oil showed good antioxidant activity, IC₅₀ of standard ascorbic acid was 30.09 µg/ml whereas that of oil was 51.17 µg/ml. In superoxide anionic scavenging method,

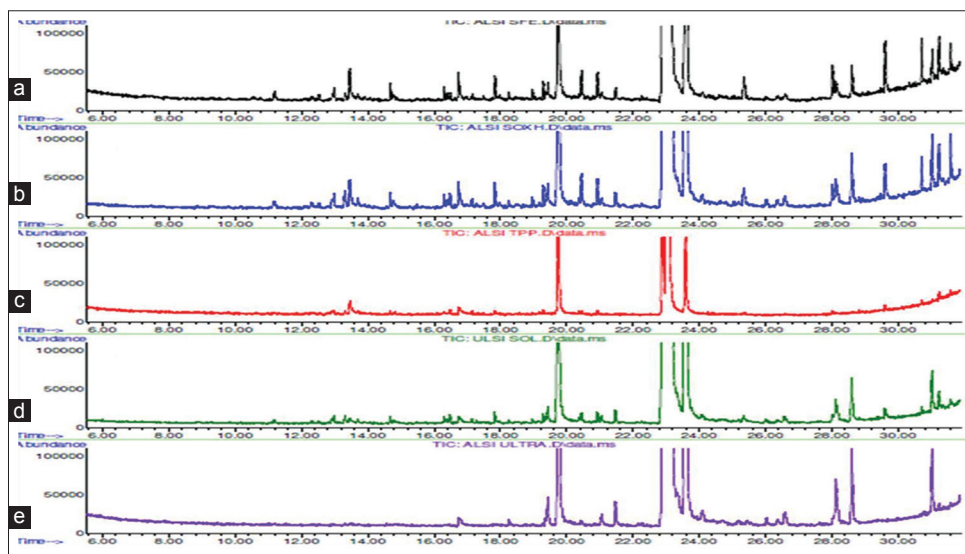


Figure 2: Gas chromatography-mass spectrometry chromatograms of fatty acid methyl esters of extracted fixed oil of peanut; (a) supercritical fluid extraction, (b) soxhlet extraction, (c) three-phase partitioning method, (d) solvent extraction, (e) ultrasound-assisted extraction

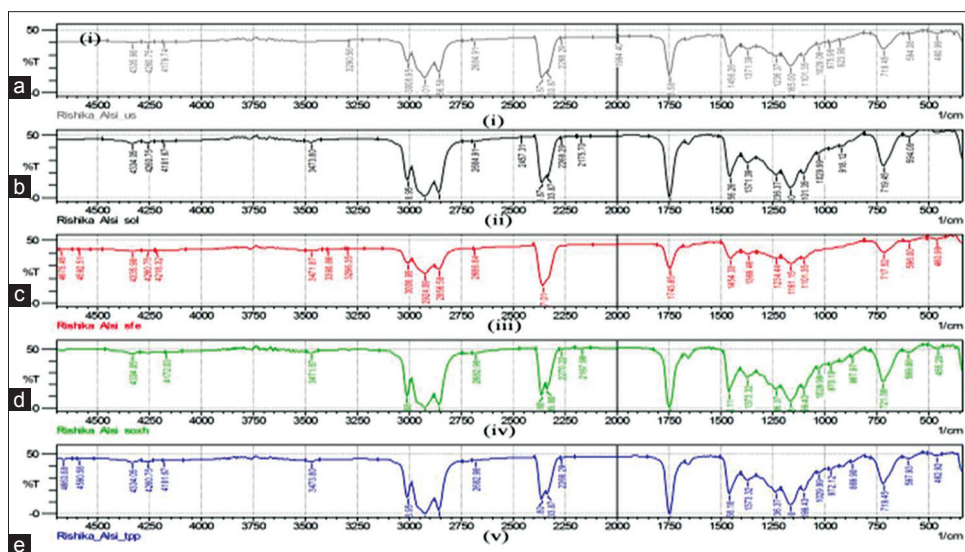


Figure 3: Comparative Fourier transform infrared spectrometry spectra of the fixed oil of peanut; (a) ultrasound-assisted extraction, (b) solvent extraction, (c) supercritical fluid extraction, (d) soxhlet extraction, (e) three-phase partitioning

the oil showed IC₅₀ of 86.34 µg/ml as compared with standard (ascorbic acid, IC₅₀ 217.40 µg/ml). The oxidative stability of peanut oil is highly correlated with the ratio of oleic acid to linoleic acid and as the ratio increases, oxidative stability increases.

Conclusion

To recover the oil of peanut, different extraction techniques were used. The SFE is a promising technique for the extraction of fixed oil. It has advantages of rapidity, low thermal damage, reduced solvent volume, and the possibility to selectively isolate the components of interest. The fatty acid profile of peanut oil revealed that oleic acid is the main monounsaturated fatty acid. The peanut oil has good antioxidant activity that is related to its lower unsaturation.

Financial support and sponsorship

Nil.

Conflicts of interest

There are no conflicts of interest.

References

1. Yoshida H, Hirakawa Y, Tomiyama Y, Mizushima Y. Effects of microwave treatment on the oxidative stability of peanut (*Arachis hypogaea*) oils and the molecular species of their triacylglycerols. *Eur J Lipid Sci Technol* 2003;105:351-8.
2. Cobb WY, Johnson BR. Physicochemical properties of peanuts. In *Peanuts: Culture and Uses*. Am Peanut Res Educ Assoc, Stillwater; 1972. p. 209-10.
3. Awad AB, Chan KC, Downie AC, Fink CS. Peanuts as a source of beta-sitosterol, a sterol with anticancer properties. *Nutr Cancer* 2000;36:238-41.
4. Sharma A, Khare SK, Gupta MN. Enzyme-assisted aqueous extraction of peanut oil. *JAACS* 2002;79:215-8.
5. Li H, Pordesimo L, Weiss J. High intensity ultrasound-assisted extraction of oil from soybeans. *Food Res Int* 2004;37:731-8.
6. Sharma A, Gupta MN. Oil extraction from almond, apricot and rice bran by three-phase partitioning after ultrasonication. *Eur J Lipid Sci Technol* 2004;106:183-6.
7. Carvalho RH, Galvao EL, Barros JA, Conceicao MM, Sousa EM. Extraction, fatty acid profile and antioxidant activity of sesame extract (*Sesamum indicum* L.). *Braz J Chem Eng* 2012;29:409-20.
8. Pharmacopoeia of India. Ministry of Health, Govt. of India 2007;1:88-90.
9. Khan R, Srivastava R, Khan MA, Alam P, Abdin MZ, Mahmooduzzafar. Variation in oil content and fatty acid composition of the seed oil of *Acacia* species collected from the Northwest zone of India. *J Sci Food Agric* 2012;92:2310-5.
10. Vlachos N, Skopelitis Y, Psaroudaki M, Konstantinidou V, Chatzilazarou A, Tegou E. Applications of Fourier transform-infrared spectroscopy to edible oils. *Anal Chim Acta* 2006;573-574:459-65.
11. Hasan MS, Ahmed MI, Mondal S, Uddin SJ, Masud MM, Sadhu SK, *et al.* Antioxidant, antinociceptive activity and general toxicity study of *Dendrophthoe falcata* and isolation of quercitrin as the major component. *Orient Pharm Exp Med* 2006;6:355-60.
12. Liu W, Fu YJ, Zu YG, Tong MH, Wu N, Liu XL, *et al.* Supercritical carbon dioxide extraction of seed oil from *Opuntia dillenii* haw and its antioxidant activity. *Food Chem* 2009;114:334-9.
13. Liu F, Ooi VE, Chang ST. Free radical scavenging activities of mushroom polysaccharide extracts. *Life Sci* 1997;60:763-71.

Author Help: Online submission of the manuscripts

Articles can be submitted online from <http://www.journalonweb.com>. For online submission, the articles should be prepared in two files (first page file and article file). Images should be submitted separately.

1) First Page File:

Prepare the title page, covering letter, acknowledgement etc. using a word processor program. All information related to your identity should be included here. Use text/rtf/doc/pdf files. Do not zip the files.

2) Article File:

The main text of the article, beginning with the Abstract to References (including tables) should be in this file. Do not include any information (such as acknowledgement, your names in page headers etc.) in this file. Use text/rtf/doc/pdf files. Do not zip the files. Limit the file size to 1 MB. Do not incorporate images in the file. If file size is large, graphs can be submitted separately as images, without their being incorporated in the article file. This will reduce the size of the file.

3) Images:

Submit good quality color images. Each image should be less than 4096 kb (4 MB) in size. The size of the image can be reduced by decreasing the actual height and width of the images (keep up to about 6 inches and up to about 1800 x 1200 pixels). JPEG is the most suitable file format. The image quality should be good enough to judge the scientific value of the image. For the purpose of printing, always retain a good quality, high resolution image. This high resolution image should be sent to the editorial office at the time of sending a revised article.

4) Legends:

Legends for the figures/images should be included at the end of the article file.